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The synthesis of two new flavin substituted calix^[4]arene derivatives, 9 and 10, is described. The first flavin substituted calix^[4]arene derivative **9** was synthesized by the reaction of 3-methylalloxazine (5) with 25,27-bis(3-bromopropoxy)-26,28-dihydroxy-5,11,17,23-tetra(tert-butyl)calix[4]arene (4) in high yield (92%). The other derivative 10 was prepared from 3-methylalloxazine-1-acetic acid (7) and 25,27 bis(3-cyanopropoxy)calix[4]arene (3). All new compounds were characterized by a combination of FT-IR and ¹H-NMR spectroscopy, and elemental-analysis techniques.

Introduction. – Flavins play an important role in living organisms [1], and they are involved in several important photobiological and photochemical processes, such as phototropism, phototaxis, and photodynamic action [2] [3]. Since 1966, the photochemistry and photophysics of alloxazine derivatives have been studied, due to the discovery of the proton-transfer reactions in lumichrome and related compounds [4] [5]. It is well-known that substituted alloxazine ($=$ benzo[g]pteridine-2,4(1H,3H)dione) derivatives, mainly lumichromes, are present in many foods and formed in the normal metabolic process of ingested riboflavin [6]. Recently, alloxazines have attracted attention due to realization of their possible involvement in a wide variety of biological systems [6] [7]. For instance, it has been shown that lumichrome may be used to inhibit flavin reductase in living Escherichia coli cells [8].

Supramolecular science has been developed tremendously over the past 30 years [9] [10]. It is known that preorganization and cooperativeness of multifunctional groups play a major role in biological reaction kinetics [11]. Some calix[4]arene derivatives, which bear azo groups [12], guadinium units [13], or nucleoside hybrids [11], are compounds which can be used as enzymes in the cell. At the same time, flavins that are obtained by condensation between α -phenylenediamine and alloxane [14] are involved in several biological processes [15]. These are generally redox cofactors involved in electron redox processes [16] of enzymatic reactions [17] [18]. To this end, we considered that compounds containing both flavin and calixarene units, *i.e.*, 9 and 10, respectively, might have interesting biological activities. Here, we report the first synthesis of calixarene derivatives that contain flavin units.

Results and Discussion. – Synthesis. The calix[4]arenes can be functionalized with desired groups both at the upper and the lower rim [19] [20] in order to obtain appropriate arrangements or to achieve the preorganized conformation [9].

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The aim of this work was the design and synthesis of new calix[4]arene derivatives, which are cited as flavin-conjugate calixarene derivatives. All new compounds were characterized by means of ¹ H-NMR and IR spectroscopy, and elemental analyses. The synthetic routes leading to new calixarene derivatives are depicted in *Schemes 1* and 2.

Scheme 1. The Synthetic Route for the Preparation of 3 and 4

i) AlCl₃, PhOH, toluene, r.t., 78%. ii) K₂CO₃, MeCN, Cl(CH₂)₃CN, reflux, 79%. iii) 1,3-Dibromopropane, K₂CO₃, MeCN, reflux, 64%.

 p -(tert-Butyl)calix[4]arene 1 was chosen as the starting material, and it was transformed to the derivatives 3 and 4 according to known procedures [19] [21] [22] (see Scheme 1). The flavin part of the molecules was synthesized starting from compound 5, which is available by condensation of benzene-1,2-diamine with N-methylalloxane according to the procedure described in $[14]$. Treatment of 5 with BrCH₂COOEt afforded ethyl 3-methylalloxazine-1-acetate (6; Scheme 2). Then, upon hydrolysis of 6 with HCl, the flavin-carboxylic acid 7 was obtained. Treatment of 7 with SOCl₂ at 45° for 2 h yielded the Cl derivative 8. The substitution of p -(tert-butyl)calix[4]arene derivative (4) at its bromoalkoxy chains was conducted in the presence of K_2CO_3 in DMF under N_2 with flavin to afford the cone conformer flavin–calix[4]arene conjugate 9 in high yield (92%). The other flavin-calix[4]arene conjugate 10 was synthesized by treatment of 1-(2-chloro-2-oxoethyl)-3-methylalloxazine (8) with 25,27-bis(3-cyanopropoxy)calix[4]arene (3) in MeCN in the presence of K_2CO_3 at 85° for 37 h in 42% yield. The $^1\mathrm{H}\text{-}\mathrm{NMR}$ spectra of $\bm{9}$ and $\bm{10}$ display a typical AX pattern for the $\mathrm{H}\text{-}$ atoms of a CH₂ bridge (ArCH₂Ar) of the calixarene moiety at $\delta(H)$ 4.17 and 3.5 ppm ($J = 12.8$) Scheme 2. The Synthetic Route for the Preparation of 9 and 10

i) BrCH₂COOEt, K₂CO₃, DMF, r.t., 77%. ii) HCl, 80 – 90°, 55%. iii) SOCl₂, 45°. iv) **4**, NaI, K₂CO₃, DMF, N_2 , r.t., 92%. $v)$ 3, MeCN, K_2CO_3 , 85°, 42%.

for 10, and 4.11 ppm $(J=12.8)$ for 9, indicating that compounds 9 and 10 exist in the cone conformation [23].

Conclusions. – We have synthesized two new calix[4]arene derivatives which contain flavin units at the lower rim of calix[4]arene. The structures of flavin calix[4] arene conjugates have been established by means of IR and 1 H-NMR spectroscopy, and elemental analyses.

It is well-known that the compounds which have N and O donor atom groups are able to inhibit the enzyme production [24]; therefore, 9 and 10, which have several donor atoms, potentially could inhibit the enzyme production, and their biological activity could be investigated in vivo and in vitro. In conclusion, 9 and 10 may be used for numerous applications in the medicinal area.

We thank the Scientific and Technical Research Council of Turkey (TUBITAK - Grant No. TBAG AY/396) for financial support of this work.

Experimental Part

General. Solvents were dried by storing them over molecular sieves (Aldrich; $4 \text{ Å}, 8-12 \text{ mesh}$). Toluene was dried with $CaH₂$ and stored over Na wire. DMF was dried with $CaSO₄$ and stored over molecular sieves. The drying agents employed during workup were Na_2SO_4 and $MgSO_4$. All aq. solns. were prepared with deionized H_2O that had been passed through a *Milli-Q Plus* water purification system. Starting materials and reagents were obtained from Aldrich, Lancaster, and Fluka and were used without further purification. TLC: DC Alufolien Kieselgel 60 F_{254} (Merck). M.p.: Barnsted/Electro thermal apparatus in a sealed capillaries; uncorrected. IR Spectra: Perkin-Elmer 1605 FTIR System Spectrum BX spectrometer; as KBr pellets. ¹H-NMR Spectra: *Varian 400* MHz and *Varian Mercury Plus* 300 MHz spectrometers; chemical shifts in ppm rel. to TMS ($\delta = 0.0$ ppm). Elemental analyses (C, H, N): Perkin-Elmer 240 analyzer.

Syntheses. 5,11,17,23-Tetra(tert-butyl)-25,26,27,28-tetrahydroxycalix[4]arene (1), 25,26,27,28-tetrahydroxycalix[4]arene (2), 25,27-bis(3-cyanopropoxy)-26,28-dihydroxycalix[4]arene (3), 25,27-bis(3-bromopropoxy)-5,11,17,23-tetra(tert-butyl)-26,28-dihydroxycalix[4]arene (4), and 3-methylalloxazine (5) were synthesized according to literature procedures [14] [19] [21] [22]. Ethyl (3-methylalloxazin-1yl)acetate (6), (3-methylalloxazin-1-yl)acetic acid (7), (3-methylalloxazin-1-yl)acetyl chloride (8), and the final products flavin-calix[4]arene 9 and flavin-calix[4]arene conjugates 9 and 10, resp., were prepared for the first time according to the procedures described below.

 $25,27$ -Bis(3-cyanopropoxy)-26,28-dihydroxycalix[4]arene (=4,4'-{[26,28-Dihydroxypentacyclo-[19.3.1.13,7.19,13.115,19]octacosa-1(25),3(28),4,6,9(27),10,12,15(26),16,18,21,23-dodecaene-25,27-diyl]bis- $(0xy)$] dibutanenitrile; 3). Prepared according to [21]. Yield 79%. M.p. 244 – 246°. IR (KBr): 2363 (CN). ${}^{1}H\text{-NMR (CDCl}_3, 400 \text{ MHz})$: 2.39 (quint., $J = 8.0, 2 \text{ CH}_2CH_2CN$); 3.08 (t, $J = 7.2, 2 \text{ CH}_2CN$); 3.47 (d, $J = 1.2, 3.4$ 12.8, 2 ArCH₂Ar); 4.13 (t, J = 5.6, 3 OCH₂CH₂); 4.21 (d, J = 12.8, 2 ArCH₂Ar); 6.70 (t, J = 7.2, 2 arom. H); 6.79 (t, $J = 7.2$, 2 arom. H); 6.94 (d, $J = 7.2$, 4 arom. H); 7.09 (d, $J = 7.2$, 4 arom. H); 7.79 (s, 2 OH). Anal. calc. for C₄H₃₄N₂O₄ (558.68): C 77.40, H 6.13, N 5.01; found: C 77.01, H 6.22, N 4.92.

 $25,27$ -Bis(3-bromopropoxy)-5,11,17,23-tetra(tert-butyl)-26,28-dihydroxycalix[4]arene (=26,28-Bis(3-bromopropoxy)-5,11,17,23-tetra-(tert-butyl)pentacyclo[19.3.1.13,7.19,13.115,19]octacosa-1(25),3(28),4, 6,9(27),10,12,15(26),16,18,21,23-dodecaene-25,27-diol; 4). Prepared according to [22]. Yield 64%. M.p. 277 – 279°. ¹H-NMR (CDCl₃, 300 MHz): 7.05 (s, 4 arom. H); 4.32 (d, J = 13.3, ArCH₂Ar); 4.28 (t, J = 5.6, 2 CH₂O); 3.83 (t, J = 6.4, 2 CH₂Br); 3.32 (d, J = 13.3, 2 ArCH₂Ar); 1.29 (s, 2 t-Bu); 0.94 (s, 2 t-Bu). Anal. calc. for $C_{50}H_{66}Br_2O_4$ (890.88): C 67.41, H 7.47, Br 17.94; found: C 67.53, H 7.50, Br 17.93.

3-Methylalloxazine $(= 3$ -Methylbenzo[g]pteridine-2,4(1H,3H)-dione; 5). Prepared according to [14]. Yield: 65%. M.p. 285°. ¹H-NMR ((D₆)DMSO, 300 MHz): 3.29 (*s*, MeN); 7.74 – 7.79 (*m*, 1 arom. H); 7.92 (d, $J = 3.5$, 2 arom. H); 8.18 (d, $J = 8.2$, 1 arom. H); 12.23 (s, NH). Anal. calc. for C₁₁H₈N₄O₂ (228.21): C 55.77, H 3.81, N 23.65; found: C 56.12, H 3.41, N 23.55.

Ethyl (3-Methylalloxazin-1-yl)acetate $(=Ethyl$ (3-Methyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin- $1(2H)-yI$ acetate; 6). A mixture of 5 (0.3 g, 1.315 mmol), K_2CO_3 (1 g, 7.303 mmol), and BrCH₂COOEt $(2.195 g, 13.146 mmol)$ were stirred for 5 h in DMF $(110 ml)$ at r.t. Then, the solvent was removed in vacuo, and the yellow product was crystallized from CH_2Cl_2/Et_2O . Yield: 0.318 g (77%). M.p. 205 – 207°. ${}^{1}H\text{-NMR } ((D_6)DMSO, 400 MHz): 1.31 (t, J=6.9, Me); 3.62 (s, MeN); 4.26 (q, J=7.5, CH_2O); 5.20 (s,$ CH₂N); 7.77 (t, J = 7.8, 1 arom. H); 7.89 (t, J = 6.8, 1 arom. H); 7.99 (d, J = 7.7, 1 arom. H); 8.36 (d, J = 8.2, 1 arom. H). Anal. calc. for C₁₅H₁₄N₄O₄ (314.3): C 57.32, H 4.49, N 17.83; found: C 57.57, H 4.32, N 17.78.

 $(3-Methylalloxazin-1-yl)$ acetic Acid $(=(3-Methyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-1(2H)-yl)$ acetic Acid; 7). A mixture of 6 (0.318 g, 1.012 mmol) and HCl (5 ml) was stirred at $80-90^\circ$ for 105 min. The mixture was cooled, and ice-water (15 ml) was added to the soln. The yellow solid that precipitated was filtered off and washed with H₂O. Reprecipitation from 2N AcOH gave 7 as a light yellow solid.

 $\rm{Yield:}\ 0.158$ g (55%). M.p. 289–293°. $\rm{^1H\text{-}NMR}$ (($\rm{D_6)DMSO},$ 400 MHz): 3.31 (s, MeN); 4.98 (s, CH₂N); 7.85 (t, $J = 6.3$, 1 arom. H); 8.00 (d, $J = 5.1$, 2 arom. H); 8.26 (d, $J = 7.8$, 1 arom. H); 13.15 (s, OH). Anal. calc. for $C_{13}H_{10}N_4O_4$ (286.24): C 54.55, H 3.52, N 19.57; found: C 54.67, H 3.45, N 19.53.

 $(3-Methylaluoxazin-1-yl)$ acetyl Chloride $(=(3-Methyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-1)$ $1(2H)-y*l*)*acetyl Chloride*; 8). Compound 7 (1.00 g, 0.350 mmol) was added to SOCl₂ (2.5 ml,$ 31.903 mmol), and the suspension was stirred at 45°. After 2 h, the solid had dissolved, $S O Cl₂$ was evaporated at $\langle 45^\circ \rangle$, and the crude chloride 8 obtained was used without further purification.

1,1'-{[5,11,17,23-Tetra-(tert-butyl)-26,28-dihydroxypentacyclo[19.3.1.13,7.19,13.115,19]octacosa-1(25), 3(28),4,6,9(27),10,12,15(26),16,18,21,23-dodecaene-25,27-diyl]bis(oxypropane-3,1-diyl)}bis(3-methyl $benzo[g]$ pteridine-2,4(IH,3H)-dione); 9). K₂CO₃ (0.6 g), 5 (0.438 mmol), and NaI (0.4 g) were added to a soln. of 4 (0.219 mmol) in 20 ml of dry DMF, and the mixture was stirred under N₂ at r.t. for 47 h. The reaction was monitored by TLC. The salts were filtered off, and the solvent from the filtrate was removed under reduced pressure. Then, $Et₂O$ was added, the precipitates formed were filtered off, and the received product was dried in a vacuum desiccator. Yield 92%. M.p. $> 350^{\circ}$. ¹H-NMR ((D₆)DMSO, 400 MHz): 8.53 (s, 2 OH); 8.07 – 7.93 (m, 4 arom. H); 7.76 – 7.76 (m, 2 arom. H); 7.58 – 7.48 (m, 2 arom. H); 7.10 (br. s, 8 arom. H); 4.62 (t, J = 6.4, 2 CH₂O); 4.11 (d, J = 12.8, 2 ArCH₂Ar); 4.0 (t, J = 5.2, 2 CH₂N); 3.35 (overlapped with DMSO, this area should correspond to 10 H-atoms belonging to ArCH₂Ar and MeN); 2.32 – 2.29 (m, 2 CH₂); 1.16 (s, 2 t-Bu); 1.10 (s, 2 t-Bu). Anal. calc. for C₇₂H₈₀N₈O₈ (1185.45); C 72.95, H 6.80, N 9.45; found: C 73.02, H 6.88, N 9.32.

26,28-Bis(3-cyanopropoxy)pentacyclo[19.3.1.1^{3,7}.19,¹³,1^{5,19}]octacosa-1(25),3(28),4,6,9(27),10,12, 15(26),16,18,21,23-dodecaene-25,27-diyl Bis[(3-methyl-2,4-dioxo-3,4-dihydrobenzo[g]pteridin-1(2H) yl)acetate]; **10**). A mixture of 8 (0.11 g, 0.36 mmol), 3 (0.09 g, 0.164 mmol), and K_2CO_3 (0.10 g, 0.724 mmol) in MeCN (6 ml) was stirred at 85° for 37 h. After cooling, MeCN was removed to dryness, and the residue was dissolved in CH₂Cl₂, washed with H₂O (5 \times 30 ml), dried (Na₂SO₄), and then concentrated to a third of the original volume on a rotary evaporation. The residue was then mixed with EtOH. Precipitated 10 was filtered off and dried under vacuum. Yield: 75 mg (42%). M.p. 340°. IR (KBr disk): 1740 (ester C=O). ¹H-NMR ((D₆)DMSO, 400 MHz): 2.32 (quint., J = 6.1, 2 CH₂CH₂CN); 3.10 (t, $J = 7.0$, 2 CH₂CN); 3.36 (s, 2 MeN); 3.49 (d, $J = 12.8$, 2 ArCH₂Ar); 4.07 (t, $J = 5.6$, 2 CH₂O); 4.17 (d, $J =$ 12.8, 2 ArCH₂Ar); 5.77 (s, 2 CH₂N); 6.63 (t, J = 7.5, 2 arom. H); 6.82 (t, J = 7.5, 2 arom. H); 7.08 (d, J = 7.6, 4 arom. H); 7.18 $(d, J = 7.5, 4 \text{ arom. H})$; 7.80 – 7.83 $(m, 2 \text{ arom. H})$; 8.01 $(d, J = 2.3, 2 \text{ arom. H})$; 8.24 $(d, J = 1.5, 4 \text{ arom. H})$ 7.9, 4 arom. H). Anal. calc. for $C_{62}H_{50}N_{10}O_{10}$ (1095.14): C 68.00, H 4.60, N 12.79; found: C 67.97, H 4.51, N 12.93.

REFERENCES

- [1] V. Massey, *Biochem. Soc. Trans.* **2000**, 28, 283.
- [2] M. Sikorski, D. Prukała, M. Insińska-Rak, I. Khmelinskii, D. R. Worrall, S. L. Williams, J. Hernando, J. L. Bourdelande, J. Koput, E. Sikorska, J. Photochem. Photobiol., A 2008, 200, 148.
- [3] H. I. Ali, N. Ashida, T. Nagamatsu, Bioorg. Med. Chem. 2008, 16, 922.
- [4] J. Koziol, Photochem. Photobiol. 1966, 5, 41.
- [5] E. Sikorska, I. V. Khmelinskii, S. L. Williams, D. R. Worrall, J. R. Herance, J. L. Bourdelande, J. Koput, M. Sikorski, J. Mol. Struct. 2004, 697, 199.
- [6] J. Chastain, D. B. McCormick, 'Flavin Metabolites', in 'Chemistry and Biochemistry of Flavoenzymes, Ed. F. Muller, CRC Press, Boston, 1991, p. 196.
- [7] P. F. Heelis, Chem. Soc. Rev. 1982, 11, 15.
- [8] O. Cunningham, M. G. Gore, T. J. Mantle, *Biochem. J.* **2000**, 345, 393.
- [9] C.-C. Zeng, Q.-Y. Zheng, Y.-L. Tang, Z.-T. Huang, Tetrahedron 2003, 59, 2539.
- [10] S. Sayin, F. Őzcan, M. Yilmaz, J. Hazard. Mater. 2010, 178, 312.
- [11] S. J. Kim, B. H. Kim, Tetrahedron Lett. 2002, 43, 6367.
- [12] T. Haino, M. Nakamura, N. Kato, M. Hiraoka, Y. Fukazawa, Tetrahedron Lett. 2004, 45, 2281.
- [13] F. Sansone, M. Dudič, G. Donofrio, C. Rivetti, L. Baldini, A. Casnati, S. Cellai, R. Ungaro, J. Am. Chem. Soc. 2006, 128, 14528.
- [14] K. Bergstad, J.-E. Bäckvall, J. Org. Chem. 1998, 63, 6650.
- [15] A. J. Robak, B. P. Branchaud, Tetrahedron Lett. 2005, 46, 5651.
- [16] R. Cibulká, L. Baxová, H. Dvořáková, F. Hampl, P. Ménová, V. Mojr, B. Plancq, S. Sayin, Collect. Czech. Chem. Commun. 2009, 74, 973.
- [17] R. B. Silverman, 'The organic chemistry of enzyme-catalyzed reactions', Academic Press, London, 2002, pp. 119 – 146.
- [18] J. McMurray, T. Begley, 'The organic chemistry of biological pathways', Roberts and Company, Colorado, 2005, pp. 109 – 111.
- [19] C. D. Gutsche, M. Iqbal, Org. Synth. 1990, 68, 234.
- [20] C. D. Gutsche, M. Iqbal, D. Stewart, J. Org. Chem. 1986, 51, 742.
- [21] E. M. Collins, M. A. McKervey, E. Madigan, M. B. Moran, M. Owens, G. Ferguson, S. J. Harris, J. Chem. Soc., Perkin Trans. 1 1991, 3137.
- [22] Z.-T. Li, G.-Z. Ji, C.-X. Zhao, S.-D. Yuan, H. Ding, C. Huang, A.-L. Du, M. Wei, J. Org. Chem. 1999, 64, 3572.
- [23] C. Jaime, X. de Mendoza, P. Prados, P. M. Nieto, C. Sanchez, J. Org. Chem. 1991, 56, 3372.
- [24] M. A. Kőroğlu, I. Erol, E. Korcan, M. Konuk, J. Macromol. Sci. Part A: Pure Appl. Chem. 2007, 44, 817.

Received May 5, 2010